(AN-FB-Architect c4000/c8000/c16000 .EN rev 2025.02.20)



#### **General Information**

#### Intended use:

FOB Turbilatex® Combo is a latex turbidimetric assay for the quantitative detection of haemoglobin (faecal occult blood) in human stool samples.

This assay is simple and widely applicable. This product is optimized for several automated analyser.

For professional in vitro diagnostic use only.

#### Reagents:

#### Materials provided by CerTest Biotec:

Reagents	Code				
Turbidimetric reagents (R1 &	TL-022FB01				
R2)	TL-022FB02				
Auxiliary Reagents					
	TL-022FB70,				
	TL-022FB71				
	TL-022FB72				
Calibration kit	TL-022FB73				
	TL-022FB74				
	TL-022FB75				
	TL-022FB08				
Controls kit	TL-022FB09				
Sample dilutions vials	MST-0019U				

#### Preparation of reagents: .

R1 and R2 are ready to use. Calibrators are ready to use.

Controls are ready to use

#### Storage and stability:

Kit components must be stored at temperature indicated on the label. Do not freeze.

Reagents are stable up to the expiration date printed on the label, always considering that reagent containers must be properly closed to avoid any contamination, must be kept away from the sunlight and conserved at temperature indicated on the label of each reagent.

#### Specimen:

Collect enough quantity of human stool samples. These samples should be collected in clean and dry containers (no preservatives or transport media). The samples can be stored in the refrigerator (2-8°C) for 3 days prior to testing. Homogenise stool samples as thoroughly as possible prior to preparation.

The sample dilution vial with diluted sample can be stored for 7 days in the refrigerator (2-8°C) prior to testing.

Use **FOB Turbilatex**® **Combo** stool collection tubes for sample collections described the instructions for use.

#### Assay procedure

FOB Turbilatex® Combo can be performed on every open chemistry analyser. Please follow the subsequent instructions in order to assure performance characteristics as describes in the instructions for use. This instruction has been validated by CerTest BIOTEC S.L Laboratories.

Additionally, please read the "Instructions for use" for instructions on operating and programming user defined test.

#### Application parameter set up:

Specific analyzers settings for **FOB Turbilatex**® **Combo** must be programmed onto the analyzer, see below. For instructions, consult the Architect c4000/c8000/c16000 (Abbott) analyzer manual and instructions for use provided with the kit.

#### Loading of reagents:

Load reagents according to the Architect c4000/c8000/c16000 (Abbott) analyzer manual.

#### **Calibration curve establishment:**

A 6-points calibration curve can be established in Architect c4000/c8000/c16000 (Abbott) analyzer. For instructions consult analyzer manual.

#### **Calibration stability:**

Calibrate the system at least once a week is extremely recommended. Recalibrate the system when reagent lot is change or when the controls are out of the assigned range given in the control label and CoA.

## QC controls:

FOB Turbilatex® Combo controls C1 and C2 must be assayed each day before running patient fecal sample extract to validate the calibration curve. The controls have assigned value ranges indicated on the label and certificate of analysis supplied. The control measurements must be within the indicated value range to obtain valid results for patient fecal extract. If the control values are out of range, follow next procedures: 1) Repeat QC control measurement, 2) Repeat calibration measurement.

#### **Results:**

The results are evaluated automatically by the analyzer and presented in ng/mL.

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#### Performance characteristics (\*)

The following results have been obtained during the validation of **FOB Turbilatex**® **Combo** on the Architect c4000/c8000/c16000 (Abbott) analyzer.

#### Linearity:

**FOB Turbilatex® Combo** using calibrator kit is linear in the calibration range of 40-1000 ng/mL.

## Measuring range:

FOB Turbilatex® Combo assay measuring range is 34.60-1953 ng hHb/mL. Samples more concentrated than 1000 ng hHb/mL must be diluted for proper quantification by the user, using additional sample buffer.

#### Prozone effect:

Studies have been made up to a concentration of 1 mg of hHb/mL of stool and no false negative results have been observed. Studies using higher concentrations have not been carried out.

Samples with concentrations up to 1953 ng hHb/mL can be measured without inhibitory prozone effect.

#### **Detection limit:**

Limit of detection (LOD): 32.33 ng hHb/mL.

Limit of quantification (LOQ): 34.60 ng hHb/mL. The lower limit of quantification is defined as the lowest actual amount of analysis that can be reliably detected. The Upper Limit of Quantification, it has not been determined since the LOQ experiment has proved that there is good quantification up to the 1000 ng hHb/mL point, which is the maximum point of the FOB Turbilatex® Combo calibration curve. This point can be quantified with a coefficient of variation lower than the %CV goal (13%).

#### Precision:

Within-laboratory and repeatability were determined according to CLSI EP05 using a standardised study design of 80 replicates per sample were evaluated (5 days  $\times$  4 runs  $\times$  4 replicates) and with an acceptance criterion of 20% CV.

	N	Mean	Repeatability		Total	
Sample		N	N (ng/mL)	Sd	CV%	Sd
1	80	50.37	4.83	9.6%	5.62	11.15%
2	80	124.40	13.57	10.8%	18.62	14.87%
3	80	232.78	3.46	1.5%	11.46	4.92%
4	80	320.80	3.97	1.2%	18.44	5.75%
5	80	403.55	6.57	1.6%	43.52	10.78%
6	80	561.96	8.78	1.6%	13.16	2.34%
7	80	992.05	50.49	5.1%	51.22	5.16%

Similarly, following the CLSI EP05 recommendations for reproducibility analysis, 80 replicates per sample were evaluated with three lots (5 days  $\times$  4 runs  $\times$  4 replicates) and with an acceptance criterion of 20% CV.

Sample	ample N Mean		Repeatability		Within lot		Reproducibility	
Sumple	(ng/mL)	(ng/mL)	Sd	CV%	Sd	CV%	Sd	CV%
1	80	49.466	4.067	8.2%	7.111	14.4%	7.111	14.4%
2	80	124.443	8.666	7.0%	16.788	13.5%	17.241	13.9%
3	80	238.002	9.261	3.9%	15.706	6.6%	15.868	6.7%
4	80	323.423	2.936	0.9%	17.057	5.3%	17.372	5.4%
5	80	411.280	4.693	1.1%	39.011	9.5%	39.180	9.5%
6	80	590.760	6.462	1.1%	29.908	5.1%	48.175	8.2%
7	80	1011.871	33.099	3.3%	43.607	4.3%	45.049	4.5%

## **Method comparison**

An evaluation was performed comparing **FOB Turbilatex® Combo** against another commercially available turbidimetric assay, which was considered as a gold standard. This evaluation has undergone with cut-off 10 µg of hHb/g of stool:

FOB Turbilatex® vs Evaluation criteria				
	Mean Value	95% confidence interval		
Sensitivity	87.5%	71.0-96.5%		
Specificity	97.9%	88.7-99.9%		
PPV	96.6%	82.2-99.9%		
NPV	92.0%	80.8-97.8%		
LR+	41.13	5.890-287.2		
LR-	0.128	0.051-0.320		

## **Shipping damage**

Please notify your distributor, if this product was received damaged.

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# Symbols key

IVD	For in vitro diagnostic use only	*	Keep dry
Ţi	Consult instructions for use		Temperature limitation
REF	Catalogue number	LOT	Lot number
	Use by	***	Manufacturer
$\sum_{n}$	Contains sufficient for <n> test</n>	DIL	Sample diluent
紫	Keep out of the sunlight		

## Manufacturer

## **CERTEST BIOTEC**

Pol. Industrial Río Gállego II,Calle J, N° 1, 50840, San Mateo de Gállego, Zaragoza (SPAIN) www.certest.es

## **NOTES**

Please refer to the instructions for use for the detailed information about the test on the following:

Synthesis; Principle; Precautions; Reagents; Specimen collection; Interpretation of results.

(\*) Analytical performance data were obtained with the Biolis 24 i(Tokyo Boeki) analyser.

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# Architect c4000/c8000/c16000, Abbott/ Application parameters

Std. No         6           R1         220 μL+ 11.8 μL (4% over suction+ 3 μL dead volume)           Sample         10 μL           R2         20 μL+ 3.8 μL (4% over suction+ 3 μL dead volume)           Others         Dispense type 2           Reaction mode         End point           Primary wavelength         505 nm           Secondary wavelength         None           Direction         Increase           Reagent blank lecture (cycle)         19 cycle           Final lecture (cycle)         30-32 cycle           Reaction time         close to 10 min           Linear range         40-1000 ng/mL           CALIBRATION           Calibration Method         Linear           Calibration set         5 calibrators + Blank           Blank         Calibrator 1 (50 ng/mL)           Calibrator 2         Calibrator 2 (200 ng/mL)           Calibrator 2         Calibrator 2 (200 ng/mL)           Calibrator 3         Calibrator 3 (250 ng/mL)           Calibrator 4         Calibrator 5 (1000 ng/mL)           Calibrator 5         Calibrator 5 (1000 ng/mL)           STEPS           Addition R1           Addition R1         Reaction R1	ASSAY PARAMETERS	
Sample         10 μL           R2         20 μL+ 3.8 μL (4% over suction+ 3 μL dead volume)           Others         Dispense type 2           Reaction mode         End point           Primary wavelength         505 nm           Secondary wavelength         None           Direction         Increase           Reagent blank lecture (cycle)         19 cycle           Final lecture (cycle)         30-32 cycle           Reaction time         close to 10 min           Linear range         40-1000 ng/mL           CALIBRATION           Calibration Method         Linear           Calibration Method         Calibration Scalibrator 9 (0 ng/mL)           Calibrator 1         Calibrator 1 (50 ng/mL)           Calibrator 2         Calibrator 1 (50 ng/mL)           Calibrator 2         Calibrator 2 (200 ng/mL)           Calibrator 3         Calibrator 3 (250 ng/mL)           Calibrator 4         Calibrator 5 (1000 ng/mL)           STEPS           Addition R1         Addition Sample           Incubation R1+S         120-180 s           Addition R2         Blank Lecture         Cycle 19           Blonk Lecture         Cycle 19	Std. No	6
R2         20 μL+ 3.8 μL (4% over suction+ 3 μL dead volume)           Others         Dispense type 2           Reaction mode         End point           Primary wavelength         505 nm           Secondary wavelength         None           Direction         Increase           Reagent blank lecture (cycle)         19 cycle           Final lecture (cycle)         30-32 cycle           Reaction time         close to 10 min           Linear range         40-1000 ng/mL           CALIBRATION           Calibration Method         Linear           Calibration set         5 calibrators + Blank           Blank         Calibrator 1 (50 ng/mL)           Calibrator 1         Calibrator 2 (alibrator 1 (50 ng/mL)           Calibrator 2         Calibrator 2 (100 ng/mL)           Calibrator 3         Calibrator 3 (250 ng/mL)           Calibrator 4         Calibrator 5 (1000 ng/mL)           STEPS           Addition R1           Addition Sample           Incubation R1+S         120-180 s           Addition R2           Blank Lecture         Cycle 19           Incubation reaction         Close to 300 sec	R1	220 μL+ 11.8 μL (4% over suction+ 3 μL dead volume)
Others         Dispense type 2           Reaction mode         End point           Primary wavelength         505 nm           Secondary wavelength         None           Direction         Increase           Reagent blank lecture (cycle)         19 cycle           Final lecture (cycle)         30-32 cycle           Reaction time         close to 10 min           Linear range         40-1000 ng/mL           CALIBRATION           Calibration Method         Linear           Calibration set         5 calibrators + Blank           Blank         Calibrator 2 (and instance)           Calibrator 1         Calibrator 2 (and instance)           Calibrator 2         Calibrator 2 (100 ng/mL)           Calibrator 3         Calibrator 3 (250 ng/mL)           Calibrator 4         Calibrator 4 (500 ng/mL)           STEPS           Addition R1         Addition Sample           Incubation R1+S         120-180 s           Addition R2         Blank Lecture         Cycle 19           Blonk Lecture         Close to 300 sec	Sample	10 μL
Reaction mode         End point           Primary wavelength         505 nm           Secondary wavelength         None           Direction         Increase           Reagent blank lecture (cycle)         19 cycle           Final lecture (cycle)         30-32 cycle           Reaction time         close to 10 min           Linear range         40-1000 ng/mL           CALIBRATION           Calibration Method         Linear           Calibration set         5 calibrators + Blank           Blank         Calibrator 0 (0 ng/mL)           Collibrator 1         Calibrator 1 (50 ng/mL)           Collibrator 2         Calibrator 2 (100 ng/mL)           Collibrator 3         Calibrator 3 (250 ng/mL)           Collibrator 4         Calibrator 4 (500 ng/mL)           STEPS           Addition R1         Addition Sample           Incubation R1+S         120-180 s           Addition R2         Blank Lecture         Cycle 19           Blonk Lecture         Close to 300 sec	R2	20 μL+ 3.8 μL (4% over suction+ 3 μL dead volume)
Primary wavelength Secondary wavelength None Direction D	Others	Dispense type 2
Secondary wavelength Direction Direction Direction Reagent blank lecture (cycle) Final lecture (cycle) Reaction time Close to 10 min Linear range A0-1000 ng/mL  CALIBRATION Calibration Method Calibration set Scalibrators + Blank Blank Calibrator 1 Calibrator 1 Calibrator 1 Calibrator 2 Calibrator 2 Calibrator 3 Calibrator 3 Calibrator 3 Calibrator 4 Calibrator 4 Calibrator 5 Calibrator 5 Calibrator 5 Calibrator 5 Calibrator 5 Calibrator 8 Calibrator 9 Calibrator 1 Calibrator 1 Calibrator 1 Calibrator 1 Calibrator 1 Calibrator 2 Calibrator 2 Calibrator 3 Calibrator 3 Calibrator 3 Calibrator 3 Calibrator 4 Calibrator 4 Calibrator 5 Cali	Reaction mode	End point
Direction         Increase           Reagent blank lecture (cycle)         19 cycle           Final lecture (cycle)         30-32 cycle           Reaction time         close to 10 min           Linear range         40-1000 ng/mL           CALIBRATION           Calibration Method         Linear           Calibration set         5 calibrators + Blank           Blank         Calibrator 1 (50 ng/mL)           Calibrator 1         Calibrator 1 (50 ng/mL)           Calibrator 2         Calibrator 2 (100 ng/mL)           Calibrator 3         Calibrator 3 (250 ng/mL)           Calibrator 4         Calibrator 4 (500 ng/mL)           Calibrator 5         Calibrator 5 (1000 ng/mL)           STEPS           Addition R1         Addition Sample           Incubation R1+S         120-180 s           Addition R2         Blank Lecture         Cycle 19           Blank Lecture         Cycle 19           Incubation reaction         close to 300 sec	Primary wavelength	505 nm
Reagent blank lecture (cycle) Final lecture (cycle) Reaction time Close to 10 min Linear range Au-1000 ng/mL  CALIBRATION  Calibration Method Linear Calibrator set Blank Calibrator 1 Calibrator 1 Calibrator 2 Calibrator 2 Calibrator 2 Calibrator 3 Calibrator 3 Calibrator 3 Calibrator 4 Calibrator 4 Calibrator 4 Calibrator 5 Calibrator 6 Calibrator 7 Calibrator 8 Calibrator 8 Calibrator 9 Calibrator 1 Calibrator 1 Calibrator 1 Calibrator 3 Calibrator 3 Calibrator 3 Calibrator 3 Calibrator 4 Calibrator 5 Cali	Secondary wavelength	None
Final lecture (cycle)         30-32 cycle           Reaction time         close to 10 min           Linear range         40-1000 ng/mL           CALIBRATION           Calibration Method         Linear           Calibration set         5 calibrators + Blank           Blank         Calibrator 0 (0 ng/mL)           Calibrator 1         Calibrator 1 (50 ng/mL)           Calibrator 2         Calibrator 2 (100 ng/mL)           Calibrator 3         Calibrator 3 (250 ng/mL)           Calibrator 4         Calibrator 4 (500 ng/mL)           Calibrator 5         Calibrator 5 (1000 ng/mL)           STEPS           Addition R1         Addition Sample           Incubation R1+S         120-180 s           Addition R2         Calibrator 5 (1000 ng/mL)           Blank Lecture         Cycle 19           Incubation reaction         close to 300 sec	Direction	Increase
Reaction time close to 10 min Linear range 40-1000 ng/mL  CALIBRATION  Calibration Method Linear Calibration set 5 calibrators + Blank Blank Calibrator 0 (0 ng/mL) Calibrator 1 Calibrator 1 (50 ng/mL) Calibrator 2 Calibrator 2 (100 ng/mL) Calibrator 3 Calibrator 3 (250 ng/mL) Calibrator 4 Calibrator 4 (500 ng/mL) Calibrator 5 Calibrator 5 (1000 ng/mL)  STEPS  Addition R1 Addition Sample Incubation R1+S 120-180 s Addition R2 Blank Lecture Cycle 19 Incubation reaction	Reagent blank lecture (cycle)	19 cycle
Linear range CALIBRATION  Calibration Method Calibration set Scalibrators + Blank Blank Calibrator 1 Calibrator 1 Calibrator 2 Calibrator 2 Calibrator 2 Calibrator 3 Calibrator 3 Calibrator 3 Calibrator 3 Calibrator 4 Calibrator 4 Calibrator 5 Calibrator 5 Calibrator 5 Calibrator 5 Calibrator 5 Calibrator 6 Calibrator 7 Calibrator 8 Calibrator 8 Calibrator 9 Calibrator 9 Calibrator 1 Calibrator 9 Calibrator 1 Calibrator 1 Calibrator 1 Calibrator 3 Calibrator 3 Calibrator 3 Calibrator 3 Calibrator 4 Calibrator 5 Calibrator 1 Calibrator 2 Calibrator 2 Calibrator 2 Calibrator 3 Calibrator 2 Calibrator 2 Calibrator 3 Calibrat	Final lecture (cycle)	30-32 cycle
CALIBRATION  Calibration Method Linear Calibration set 5 calibrators + Blank Blank Calibrator 1 (50 ng/mL) Calibrator 1 Calibrator 2 (100 ng/mL) Calibrator 2 Calibrator 2 (100 ng/mL) Calibrator 3 Calibrator 3 (250 ng/mL) Calibrator 4 Calibrator 4 (500 ng/mL) Calibrator 5 Calibrator 5 (1000 ng/mL) STEPS  Addition R1 Addition Sample Incubation R1+S 120-180 s Addition R2 Blank Lecture Cycle 19 Incubation reaction close to 300 sec	Reaction time	close to 10 min
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Calibration set  Blank Calibrator 0 (0 ng/mL) Calibrator 1 Calibrator 2 Calibrator 2 Calibrator 3 Calibrator 3 Calibrator 3 Calibrator 3 Calibrator 4 Calibrator 4 Calibrator 5 Calibrator 5 Calibrator 5 Calibrator 5 Calibrator 5 Calibrator 6 Calibrator 7 Calibrator 8 Calibrator 8 Calibrator 9 Calibrator 1 Calibrator 1 Calibrator 1 Calibrator 2 Calibrator 3 Calibrator 5 Calibrator 1 Calibrator 1 Calibrator 1 Calibrator 1 Calibrator 3 Calibrator	CALIBRATION	'
Blank Calibrator 0 (0 ng/mL) Calibrator 1 Calibrator 0 (0 ng/mL) Calibrator 2 Calibrator 2 (100 ng/mL) Calibrator 3 Calibrator 3 (250 ng/mL) Calibrator 4 Calibrator 4 (500 ng/mL) Calibrator 5 Calibrator 5 (1000 ng/mL)  STEPS  Addition R1 Addition Sample Incubation R1+S 120-180 s Addition R2 Blank Lecture Cycle 19 Incubation reaction	Calibration Method	Linear
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Calibrator 2 Calibrator 2 (100 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 4 (500 ng/mL) Calibrator 5 (200 ng/mL) Calibrator 5 (200 ng/mL)  STEPS  Addition R1 Addition Sample Incubation R1+S Addition R2 Blank Lecture Cycle 19 Incubation reaction Calibrator 2 (100 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 4 (500 ng/mL) Calibrator 5 (1000 ng/mL)	Blank	Calibrator 0 (0 ng/mL)
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Calibrator 4 Calibrator 4 (500 ng/mL) Calibrator 5 Calibrator 5 (1000 ng/mL)  STEPS  Addition R1 Addition Sample Incubation R1+S Addition R2 Blank Lecture Incubation reaction Calibrator 4 (500 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 6 (1000 ng/mL) Calibrator 6 (1000 ng/mL) Calibrator 7 (1000 ng/mL) Calibrator 7 (1000 ng/mL) Calibrator 7 (1000 ng/mL) Calibrator 8 (1000 ng/mL) Calibrator	Calibrator 2	Calibrator 2 (100 ng/mL)
Calibrator 5 (1000 ng/mL)  STEPS  Addition R1  Addition Sample Incubation R1+S  Addition R2  Blank Lecture Incubation reaction  Calibrator 5 (1000 ng/mL)	Calibrator 3	Calibrator 3 (250 ng/mL)
STEPS  Addition R1  Addition Sample Incubation R1+S  Addition R2  Blank Lecture Incubation reaction  Cycle 19 Incubation reaction	Calibrator 4	Calibrator 4 (500 ng/mL)
Addition R1 Addition Sample Incubation R1+S Addition R2 Blank Lecture Incubation reaction  Cycle 19 Incubation reaction	Calibrator 5	Calibrator 5 (1000 ng/mL)
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Incubation R1+S  Addition R2  Blank Lecture Incubation reaction  120-180 s  Cycle 19  Cycle 19	Addition R1	
Addition R2  Blank Lecture Cycle 19 Incubation reaction close to 300 sec	Addition Sample	
Blank Lecture Cycle 19 Incubation reaction close to 300 sec	Incubation R1+S	120-180 s
Incubation reaction close to 300 sec	Addition R2	
	Blank Lecture	Cycle 19
Final lecture Cycle 30-32	Incubation reaction	close to 300 sec
	Final lecture	Cycle 30-32