(AN-FB-BS-230 .EN rev 2025.02.20)



General Information

Intended use:

FOB Turbilatex® Combo is a latex turbidimetric assay for the quantitative detection of haemoglobin (faecal occult blood) in human stool samples.

This assay is simple and widely applicable. This product is optimized for several automated analyser.

For professional in vitro diagnostic use only.

Reagents:

Materials provided by CerTest Biotec:

Reagents	Code				
Turbidimetric reagents (R1 &	TL-022FB01				
R2)	TL-022FB02				
Auxiliary Reagents					
	TL-022FB70,				
	TL-022FB71				
	TL-022FB72				
Calibration kit	TL-022FB73				
	TL-022FB74				
	TL-022FB75				
	TL-022FB08				
Controls kit	TL-022FB09				
Sample dilutions vials	MST-0019U				

Preparation of reagents: .

R1 and R2 are ready to use.

Calibrators are ready to use.

Controls are ready to use

Storage and stability:

Kit components must be stored at temperature indicated on the label. Do not freeze.

Reagents are stable up to the expiration date printed on the label, always considering that reagent containers must be properly closed to avoid any contamination, must be kept away from the sunlight and conserved at temperature indicated on the label of each reagent.

Specimen:

Collect enough quantity of human stool samples. These samples should be collected in clean and dry containers (no preservatives or transport media). The samples can be stored in the refrigerator (2-8°C) for 3 days prior to testing. Homogenise stool samples as thoroughly as possible prior to preparation.

The sample dilution vial with diluted sample can be stored for 7 days in the refrigerator (2-8°C) prior to testing.

Use **FOB Turbilatex**® **Combo** stool collection tubes for sample collections described the instructions for use.

Assay procedure

FOB Turbilatex® Combo can be performed on every open chemistry analyser. Please follow the subsequent instructions in order to assure performance characteristics as describes in the instructions for use. This instruction has been validated by CerTest BIOTEC S.L Laboratories.

Additionally, please read the "Instructions for use" for instructions on operating and programming user defined test.

Application parameter set up:

Specific analyzers settings for **FOB Turbilatex® Combo** must be programmed onto the analyzer, see below. For instructions, consult the BS-230 (Midray) analyzer manual and instructions for use provided with the kit.

Loading of reagents:

Load reagents according to the BS-230 (Midray) analyzer manual.

Calibration curve establishment:

A 6-points calibration curve can be established in BS-230 (Midray) analyzer. For instructions consult analyzer manual.

Calibration stability:

Calibrate the system at least once a week is extremely recommended. Recalibrate the system when reagent lot is change or when the controls are out of the assigned range given in the control label and CoA.

QC controls:

FOB Turbilatex® Combo controls C1 and C2 must be assayed each day before running patient fecal sample extract to validate the calibration curve. The controls have assigned value ranges indicated on the label and certificate of analysis supplied. The control measurements must be within the indicated value range to obtain valid results for patient fecal extract. If the control values are out of range, follow next procedures: 1) Repeat QC control measurement, 2) Repeat calibration measurement.

Results:

The results are evaluated automatically by the analyzer and presented in ng/mL.

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Performance characteristics (*)

The following results have been obtained during the validation of **FOB Turbilatex**® **Combo** on the BS-230 (Midray) analyzer. **Linearity:**

FOB Turbilatex® Combo using calibrator kit is linear in the calibration range of 40-1000 ng/mL.

Measuring range:

FOB Turbilatex® Combo assay measuring range is 34.60-1953 ng hHb/mL. Samples more concentrated than 1000 ng hHb/mL must be diluted for proper quantification by the user, using additional sample buffer.

Prozone effect:

Studies have been made up to a concentration of 1 mg of hHb/mL of stool and no false negative results have been observed. Studies using higher concentrations have not been carried out.

Samples with concentrations up to 1953 ng hHb/mL can be measured without inhibitory prozone effect.

Detection limit:

Limit of detection (LOD): 32.33 ng hHb/mL.

Limit of quantification (LOQ): 34.60 ng hHb/mL. The lower limit of quantification is defined as the lowest actual amount of analysis that can be reliably detected. The Upper Limit of Quantification, it has not been determined since the LOQ experiment has proved that there is good quantification up to the 1000 ng hHb/mL point, which is the maximum point of the FOB Turbilatex® Combo calibration curve. This point can be quantified with a coefficient of variation lower than the %CV goal (13%).

Precision:

Within-laboratory and repeatability were determined according to CLSI EP05 using a standardised study design of 80 replicates per sample were evaluated (5 days x 4 runs x 4 replicates) and with an acceptance criterion of 20% CV.



	Mean		Repeatability		Total	
Sample	N	(ng/mL)	Sd	CV%	Sd	CV%
1	80	50.37	4.83	9.6%	5.62	11.15%
2	80	124.40	13.57	10.8%	18.62	14.87%
3	80	232.78	3.46	1.5%	11.46	4.92%
4	80	320.80	3.97	1.2%	18.44	5.75%
5	80	403.55	6.57	1.6%	43.52	10.78%
6	80	561.96	8.78	1.6%	13.16	2.34%
7	80	992.05	50.49	5.1%	51.22	5.16%

Similarly, following the CLSI EP05 recommendations for reproducibility analysis, 80 replicates per sample were evaluated with three lots (5 days \times 4 runs \times 4 replicates) and with an acceptance criterion of 20% CV.

Sample	Sample N Mean		Repeatability		Within lot		Reproducibility	
Sumple	IN	(ng/mL)	Sd	CV%	Sd	CV%	Sd	CV%
1	80	49.466	4.067	8.2%	7.111	14.4%	7.111	14.4%
2	80	124.443	8.666	7.0%	16.788	13.5%	17.241	13.9%
3	80	238.002	9.261	3.9%	15.706	6.6%	15.868	6.7%
4	80	323.423	2.936	0.9%	17.057	5.3%	17.372	5.4%
5	80	411.280	4.693	1.1%	39.011	9.5%	39.180	9.5%
6	80	590.760	6.462	1.1%	29.908	5.1%	48.175	8.2%
7	80	1011.871	33.099	3.3%	43.607	4.3%	45.049	4.5%

Method comparison

An evaluation was performed comparing **FOB Turbilatex**® **Combo** against another commercially available turbidimetric assay, which was considered as a gold standard. This evaluation has undergone with cut-off 10 µg of hHb/g of stool:

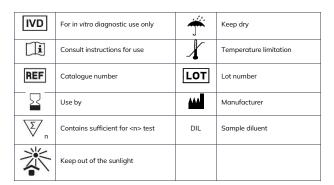
FOB Turbilatex® vs Evaluation criteria					
	Mean Value	95% confidence interval			
Sensitivity	87.5%	71.0-96.5%			
Specificity	97.9%	88.7-99.9%			
PPV	96.6%	82.2-99.9%			
NPV	92.0%	80.8-97.8%			
LR+	41.13	5.890-287.2			
LR-	0.128	0.051-0.320			

Shipping damage

Please notify your distributor, if this product was received damaged.

(AN-FB-BS-230 .EN rev 2025.02.20)

Symbols key



Manufacturer

CERTEST BIOTEC

Pol. Industrial Río Gállego II,Calle J, N° 1, 50840, San Mateo de Gállego, Zaragoza (SPAIN) www.certest.es

NOTES

Please refer to the instructions for use for the detailed information about the test on the following:

Synthesis; Principle; Precautions; Reagents; Specimen collection; Interpretation of results.

(*) Analytical performance data were obtained with the Biolis 24 i(Tokyo Boeki) analyser.



(AN-FB-BS-230 .EN rev 2025.02.20)



BS-230, Midray/ Application parameters

R1 200 µL Sample 20 µL R2 55 µL Others N/A Reaction mode End point Primary wavelength 510 nm Secondary wavelength 800 nm Direction Positive Reagent blank lecture (cycle) 2 cycle Final lecture (cycle) 20 cycle Reaction time close to 10 min Linear range 40-1000 ng/mL CALIBRATION Linear Calibration Method Linear Calibration set 5 calibrators + Blank Calibrator 1 Calibrator 2 (20 ng/mL) Calibrator 2 Calibrator 1 (50 ng/mL) Calibrator 2 Calibrator 2 (100 ng/mL) Calibrator 3 Calibrator 3 (250 ng/mL) Calibrator 4 Calibrator 4 (500 ng/mL) Calibrator 5 Calibrator 5 (1000 ng/mL) STEPS Addition R1 Addition Sample Incubation R1+S 120-180 s Addition R2 Blank Lecture Cycle 21-22 Incubation reaction	ASSAY PARAMETERS	
Sample 20 µL R2 55 µL Others N/A Reaction mode End point Primary wavelength 510 nm Secondary wavelength 800 nm Direction Positive Reagent blank lecture (cycle) 2 cycle Final lecture (cycle) 20 cycle Reaction time close to 10 min Linear range 40-1000 ng/mL CALIBRATION Calibration Method Linear Calibration set 5 calibrators + Blank Blank Calibrator 0 (on g/mL) Calibrator 1 Calibrator 1 (50 ng/mL) Calibrator 2 Calibrator 2 (100 ng/mL) Calibrator 3 Calibrator 3 (250 ng/mL) Calibrator 4 Calibrator 3 (250 ng/mL) Calibrator 5 Calibrator 5 (1000 ng/mL) STEPS Addition R1 Addition Sample Incubation R1+S 120-180 s Addition R2 Blank Lecture Cycle 21-22 Incubation reaction close to 300 sec	Std. No	6
R2 55 µL Others N/A Reaction mode End point Primary wavelength 510 nm Secondary wavelength 800 nm Direction Positive Reagent blank lecture (cycle) 2 cycle Final lecture (cycle) 20 cycle Reaction time close to 10 min Linear range 40-1000 ng/mL CAlibration Method Calibration set 5 calibrators + Blank Blank Calibrator 0 (on g/mL) Calibrator 1 Calibrator 1 (50 ng/mL) Calibrator 2 Calibrator 2 (100 ng/mL) Calibrator 3 Calibrator 3 (250 ng/mL) Calibrator 4 Calibrator 5 (200 ng/mL) Calibrator 5 Calibrator 5 (1000 ng/mL) STEPS Addition R1 Addition Sample Incubation R1+S 120-180 s Addition R2 Blank Lecture Cycle 21-22 Incubation reaction close to 300 sec	R1	200 μL
Others N/A Reaction mode End point Primary wavelength 510 nm Secondary wavelength 800 nm Direction Positive Reagent blank lecture (cycle) 2 cycle Final lecture (cycle) 20 cycle Reaction time close to 10 min Linear range 40-1000 ng/mL CALIBRATION Calibration Method Linear Calibration set 5 calibrators + Blank Blank Calibrator 0 (0 ng/mL) Calibrator 1 Calibrator 1 (50 ng/mL) Calibrator 2 Calibrator 2 (100 ng/mL) Calibrator 3 Calibrator 3 (250 ng/mL) Calibrator 4 Calibrator 5 (1000 ng/mL) STEPS Addition R1 Addition Sample Incubation R1+S 120-180 s Addition R2 Blank Lecture Cycle 21-22 Blonk Lecture Colose to 300 sec	Sample	20 μL
Reaction mode End point Primary wavelength 510 nm Secondary wavelength 800 nm Direction Positive Reagent blank lecture (cycle) 2 cycle Final lecture (cycle) 20 cycle Reaction time close to 10 min Linear range 40-1000 ng/mL CALIBRATION Calibration Method Linear Calibration set 5 calibrators + Blank Blank Calibrator 0 (0 ng/mL) Calibrator 1 Calibrator 2 (200 ng/mL) Calibrator 2 Calibrator 3 (250 ng/mL) Calibrator 3 Calibrator 3 (250 ng/mL) Calibrator 4 Calibrator 5 (1000 ng/mL) STEPS Addition R1 Addition Sample Incubation R1+S 120-180 s Addition R2 Blank Lecture Cycle 21-22 Incubation reaction close to 300 sec	R2	55 μL
Primary wavelength 510 nm Secondary wavelength 800 nm Direction Positive Reagent blank lecture (cycle) 2 cycle Final lecture (cycle) 20 cycle Reaction time close to 10 min Linear range 40-1000 ng/mL CALIBRATION Calibration Method Linear Calibration set 5 calibrators + Blank Blank Calibrator 1 (50 ng/mL) Calibrator 1 Calibrator 1 (50 ng/mL) Calibrator 2 Calibrator 2 (100 ng/mL) Calibrator 3 Calibrator 3 (250 ng/mL) Calibrator 4 Calibrator 4 (500 ng/mL) STEPS Addition R1 Addition Sample Incubation R2 Blank Lecture Cycle 21-22 Incubation reaction close to 300 sec	Others	N/A
Secondary wavelength Direction Positive Reagent blank lecture (cycle) Pinal lecture (cycle) Peaction time Positive Reaction time R	Reaction mode	End point
Direction Positive Reagent blank lecture (cycle) 2 cycle Final lecture (cycle) 20 cycle Reaction time close to 10 min Linear range 40-1000 ng/mL CALIBRATION Calibration Method Linear Calibrator set 5 calibrators + Blank Blank Calibrator 1 (50 ng/mL) Calibrator 1 Calibrator 1 (20 ng/mL) Calibrator 2 Calibrator 2 (100 ng/mL) Calibrator 3 Calibrator 3 (250 ng/mL) Calibrator 4 Calibrator 4 (500 ng/mL) STEPS Addition R1 Addition Sample Incubation R1+S 120-180 s Addition R2 Blank Lecture Cycle 1-22 Incubation reaction close to 300 sec	Primary wavelength	510 nm
Reagent blank lecture (cycle) Final lecture (cycle) Reaction time Close to 10 min Linear range CALIBRATION Calibration Method Calibrator set Blank Calibrator 1 Calibrator 1 Calibrator 1 Calibrator 2 Calibrator 2 Calibrator 3 Calibrator 3 Calibrator 3 Calibrator 4 Calibrator 4 Calibrator 5 Calibrator 5 Calibrator 5 Calibrator 5 Calibrator 5 Calibrator 5 Calibrator 8 Addition R1 Addition Sample Incubation R2 Blank Lecture Cycle 21-22 Incubation reaction	Secondary wavelength	800 nm
Final lecture (cycle) Reaction time Close to 10 min Linear range 40-1000 ng/mL CALIBRATION Calibration Method Calibration set Blank Calibrator 1 Calibrator 1 Calibrator 2 Calibrator 2 Calibrator 2 Calibrator 3 Calibrator 3 Calibrator 4 Calibrator 4 Calibrator 5 Calibrator 5 Calibrator 5 Calibrator 5 Calibrator 5 Calibrator 8 Addition R1 Addition R1 Addition R2 Blank Lecture Cycle 21-22 Incubation reaction	Direction	Positive
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Linear range CALIBRATION Calibration Method Calibration set S calibrators + Blank Blank Calibrator 1 (50 ng/mL) Calibrator 2 (100 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 4 (500 ng/mL) Calibrator 5 Calibrator 5 (1000 ng/mL) STEPS Addition R1 Addition R1 Addition R2 Blank Lecture CALIBRATION Linear Ad0-1000 ng/mL Calibrator 4 (500 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 6 (1000 ng/mL) Calibrator 7 (1000 ng/mL) Calibrator 8 (1000 ng/mL) Calibrator 9 (1000 ng/mL) Calibrator 1 (50 ng/mL) Calibrator 1 (50 ng/mL) Calibrator 2 (100 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 2 (100 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 2 (100 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 2 (100 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 2 (100 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 2 (100 ng/mL) Calibrator 2 (100 ng/mL) Calibrator 2 (100 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 2 (100 ng/mL) Calibrator 2 (100 ng/mL) Calibrator 3 (250 ng/mL) Calibrator 4 (500 ng/mL) Calibrator 4 (500 ng/mL) Calibrator 4 (500 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 6 (1000 ng/mL) Calibrator 7 (1000 ng/mL)	Final lecture (cycle)	20 cycle
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Calibration set Blank Calibrator 0 (0 ng/mL) Calibrator 1 Calibrator 1 (50 ng/mL) Calibrator 2 Calibrator 2 Calibrator 3 Calibrator 3 (250 ng/mL) Calibrator 4 Calibrator 4 Calibrator 5 Calibrator 5 (1000 ng/mL) STEPS Addition R1 Addition Sample Incubation R1+S Addition R2 Blank Lecture Cycle 21-22 Incubation reaction	CALIBRATION	
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Calibrator 3 (250 ng/mL) Calibrator 4 (500 ng/mL) Calibrator 5 (250 ng/mL) STEPS Addition R1 Addition Sample Incubation R1+S Addition R2 Blank Lecture Incubation reaction Calibrator 3 (250 ng/mL) Calibrator 4 (500 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL)	Calibrator 1	Calibrator 1 (50 ng/mL)
Calibrator 4 Calibrator 4 (500 ng/mL) Calibrator 5 Calibrator 5 (1000 ng/mL) STEPS Addition R1 Addition Sample Incubation R1+S Addition R2 Blank Lecture Incubation reaction Calibrator 4 (500 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL) Calibrator 5 (1000 ng/mL)	Calibrator 2	Calibrator 2 (100 ng/mL)
Calibrator 5 Calibrator 5 (1000 ng/mL) STEPS Addition R1 Addition Sample Incubation R1+S Addition R2 Blank Lecture Incubation reaction Calibrator 5 (1000 ng/mL)	Calibrator 3	Calibrator 3 (250 ng/mL)
STEPS Addition R1 Addition Sample Incubation R1+S Addition R2 Blank Lecture Incubation reaction Cycle 21-22 Incubation reaction	Calibrator 4	Calibrator 4 (500 ng/mL)
Addition R1 Addition Sample Incubation R1+S Addition R2 Blank Lecture Incubation reaction Cycle 21-22 Incubation reaction	Calibrator 5	Calibrator 5 (1000 ng/mL)
Addition Sample Incubation R1+S Addition R2 Blank Lecture Incubation reaction Cycle 21-22 Incubation reaction	STEPS	
Incubation R1+S Addition R2 Blank Lecture Cycle 21-22 Incubation reaction Close to 300 sec	Addition R1	
Addition R2 Blank Lecture Cycle 21-22 Incubation reaction close to 300 sec	Addition Sample	
Blank Lecture Cycle 21-22 Incubation reaction close to 300 sec	Incubation R1+S	120-180 s
Incubation reaction close to 300 sec	Addition R2	
	Blank Lecture	Cycle 21-22
Final lecture Cycle 20	Incubation reaction	close to 300 sec
	Final lecture	Cycle 20